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Postulation of leaf rust resistance genes of 20 wheat cultivars in southern Russia

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Abstract

Gene postulation is one of the fastest and most cost-effective methods for identifying seedling leaf rust resistance genes in wheat cultivars. Many researchers use this approach to identify Lr genes in wheat cultivars. The purpose of our research was to identify seedling leaf rust resistance genes in 20 wheat cultivars from different breeding centers of Russia, Ukraine and Germany. Forty-two near isogenic Thatcher lines and 10 Puccinia triticina isolates were used for gene postulation. When assessing the infection types to cultivars and lines, a scale was used, according to Oelke and Kolmer. In 20 wheat cultivars 19 Lr genes were postulated: 2c, 3, 10, 3bg, 3ka, 14a, 17, 18, 23, 25, 26, 30, 33, 40, 44, 50, B, Exch, Kanred. The most common for cultivars was the Lr10 gene. In five cultivars, showing high field resistance, most postulated seedling genes (Lr2c, Lr3, Lr10, Lr14a, Lr26, Lr33) were not effective in the adult stage. It is possible that resistance of such cultivars is associated with APR genes, the postulation of which requires an expansion in the number and spectrum of P. triticina isolate virulence. Most of the studied cultivars (60%) have recently been entered into the register (2015–2019) and in the field show a stable or moderately susceptible response to P. triticina infection, despite the fact that the Lr genes postulated in them were not effective in the adult stage. The data obtained indicated a variety of genotypes of the studied cultivars, as well as the tendency of breeders to use the effect of pyramiding ineffective genes, which can prolong the resistance of the cultivar. Annual monitoring of varieties is necessary in each region, especially when reacting with a medium susceptible type (MS), which may indicate the initial stage of resistance loss.

Keywords: Lr genes, Puccinia triticina, resistance cultivars, winter wheat

Introduction

Leaf rust (*Puccinia triticina* Erikss.) is one of the most common and harmful diseases of winter wheat in all grain-producing regions of the world. In Russia, the disease is especially common in the southern region, which is the country's leading grain producer. In this region, leaf rust is found everywhere and occurs almost annually in winter wheat crops, depending on the prevailing weather conditions (Volkova *et al.* 2019). Even though progress has been made in studying the structure and variability of leaf rust pathogen populations of the fungus *P. triticina* and the success of practical breeding for resistance, this disease leads to a loss of 15–25% of the crop yield (Sanin and Nazarova 2010). Yield losses from leaf rust are very significant even in developed countries with a high level of agriculture and chemicalization of production (Shcherbik and Kovalenko 2011).

The most economical and bio-safe method of protecting wheat from a pathogen is the cultivation of rust-resistant cultivars. It is important to know the genetics of wheat resistance. This is necessary to draw up various displacement strategies in each grain-sowing region. Gene postulation is one of the fastest methods for identifying seedling leaf rust resistance genes in winter wheat cultivars. This approach is based on

the principle of interaction of the parasite and the host according to "gene-for-gene" type (Flor 1971). The presence of resistance genes is postulated based on the expression of infectious types of wheat differentiator lines in response to infection with fungal isolates of different virulence (Wamishe et al. 2004). With a similar reaction of the isogenic line and cultivar to the defeat by most fungal isolates with different virulence, the Lr gene of the line and cultivar may coincide (Wamishe and Milus 2004). Many researchers use this approach to identify Lr genes in wheat cultivars. For example, scientists from Ethiopia and Germany tested 36 winter wheat cultivars using 31 P. triticina isolates. It was established that Lr-genes: 1, 2c, 3, 3ka, 9, 10, 14a, 14b, 13, 16, 18, 21, 23, 27 + 31, 30, 37 and 44 were postulated in Ethiopic wheat cultivars, and Lr9, Lr20 and Lr21 - in German wheat cultivars (Mebrate et al. 2008). In 66 cultivars from Argentina, using 17 different leaf rust pathotypes, 11 different genes were postulated: Lr1, Lr3a, Lr3ka, Lr9, Lr10, Lr16, Lr17, Lr19, Lr24, Lr26, and Lr4 (Vanzetti et al. 2011). This method, in combination with the use of molecular markers, can also be used to postulate the genes of adult plants (APR genes) (Wei et al. 2015; Li et al. 2018; Baidya et al. 2019). A lot of work on the postulation of seedling leaf rust resistance genes has been done in China (Li et al. 2010; Li et al. 2016; Gebrewahid et al. 2017). A similar approach, combined with the use of molecular markers, has been used by scientists from Egypt (Abouzied et al. 2017). In Russia, such studies were first conducted at the All-Russian Research Institute of Biological Plant Protection (Anpilogova et al. 2011). Seedling resistance genes to P. triticina were postulated in 18 released winter wheat cultivars. It was found that most of them were ineffective against the North Caucasian pathogen population. In 2014, we carried out a phytopathological assessment of 12 bread winter wheat cultivars using 16 pathogen isolates. Eight cultivars succeeded in postulating Lr genes: 1+, 2a, 3ka, 15, 16, 23, 33, 34 (Volkova and Vaganova 2016).

The aim of our study was to identify seedling leaf rust resistance genes in 20 wheat cultivars using 10 *P. triticina* phenotypes with different virulence.

Materials and Methods

Field studies were carried out in 2017 on the experimental field of the All-Russian Research Institute of Biological Plant Protection.

In our study we used 18 winter and two spring soft and bread wheat cultivars of foreign and Russian breeding (Table 1), from the collection of the Federal Research Center «N.I. Vavilov All-Russian Institute of Plant Genetic Resources» (VIR) as well as those of economic importance (Anisimovka, Anka, Antonina, Argonavt, Bagira, Bogdanka, Wintergold, Eremeevna, Krucha, Odari, Olkhon, Tulaykovskaya 110, Shef, Eirena, Ekada-113, Etude, Yubilyarka, Yakhont, Vidrada, Sidor Kovpak). Cultivar testing for pathogen infection was performed using 10 *P. triticina* phenotypes and 42 near isogenic Thatcher lines.

For testing wheat cultivars, 10 *P. triticina* phenotypes with different virulence were selected (Table 2). Each phenotype was assigned a four-letter code, according to the nomenclature of Long and Kolmer (Long and Kolmer 1989). Seven to nine day-old seedlings of 20 cultivars and 42 isogenic lines were separately inoculated with a spore suspension of each fungal isolate (from a spray) and placed in a humid chamber at 20°C for 16 h. Then, the inoculated plants were returned to greenhouse conditions.

The assessment was carried out on the 12th day after inoculation using the Long and Kolmer scale (17): 0 = absence of hypersensitive flecks, necrosis, or uredinia, 0; = weak hypersensitive flecks, ; = distinct hypersensitive flecks, 1 = small uredinia surrounded by distinct necrosis, 2 = small uredinia, surrounded by distinct chlorosis, 3 = moderate uredinia without chlorosis and necrosis, 4 = very large uredinia without chlorosis and necrosis. A mixture of two or more infection types (IT) was recorded as the IT prevailing first. The designations + and – are additional designations of IT from 0 to 4 and indicate larger or smaller pustules, in contrast to normal ones, respectively. DT from 0 to 2+ is understood as low, 3-4 – high.

In the field, wheat cultivars and lines were sown on plots in 6 rows 1 m long. The distance between plots was 0.5 m. In each plot there were 50-60 plants. Infection was carried out by a natural population of the fungus in the presence of drip-liquid moisture (Anpilogova and Volkova 2000). The degree of leaf rust damage and the types of reaction were taken into account 10-14 days after inoculation, with repeated counting every 10 days as urediniogenesis increased. When assessing damage types, the following scale was used, according to Oelke and Kolmer (2004): 0 = noflecks or uredinia, TR = insignificant level of uredinia, R = small uredinia with necrosis, M = a mixture of small and large uredinia with chlorosis, MR = moderate size uredinia with necrosis, MS = moderate size uredinia with chlorosis, S = large uredinia.

Results

Table 2 shows the infectious types of isogenic lines obtained by inoculation with isolates of the North Caucasian population of *P. triticina* in the seedling phase. Table 3 shows the infectious types of cultivars

winter		Country of origin	Year of entering the State Register of the Russian Federation	Originator				
		Ukraine	2012	Plant Breeding and Genetics Institute of the Ukrainian Academy of Agrarian Sciences				
Wintergold	winter durum	Germany	_	_				
Yakhont	winter durum	Russia	2018					
Etude	winter bread	Russia	2019					
Shef	winter bread	Russia	2019	Federal State Budgetary Scientific Institution Agrarian Scientific Center «Donskoy»				
Yubilyarka	winter durum	Russia	2019					
Eirena	winter durum	Russia	2017					
Olkhon	winter bread	Russia	2014					
Eremeevna	winter bread	Russia	2015					
Antonina	winter bread	Russia	2016	Federal State Budgetary Scientific Institution				
Anka	winter bread	Russia	2016	National Grain Center named after P.P. Lukyanenko				
Krucha	winter durum	Russia	2015					
Odari	winter durum	Russia	2017					
Bagira	winter bread	Russia	2013	Federal State Budgetary Scientific Institution				
Anisimovka	winter bread	Russia	-	North Caucasus Federal Agrarian Research Center				
Tulaykovskaya 110	spring bread	Russia	2015	Federal State Budgetary Scientific Institution Samara Research Scientific Institute of Agriculture named after N.M. Tulaykov				
Bogdanka	winter bread	Russia	2009	Federal State Budgetary Scientific Institution Belgorod Federal Agrarian Research Center of Russian Academy of Science				
Vidrada	winter bread	Ukraine	2011	Belotserkovskaya Experimental Breeding Station of the Institute of Sugar Beet of the Ukrainian Academy of Agrarian Science				
Sidor Kovpak	winter bread	Ukraine	-	Poltava State Agrarian Academy				
Ekada 113	spring bread	Russia	2014	Federal State Budgetary Scientific Institution Bashkir Agricultural Research Institute				

Table 1. List of winter wheat cultivars used for Lr resistance gene postulation	on
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″–″ unknown

infected with the same isolates of the North Caucasian population of *P. triticina*. The presence of seedling leaf rust resistance genes in wheat cultivars was postulated based on a comparison of their high and low ITs with infection types to isogenic lines of the Thatcher cultivar.

Seedling resistance

All studied isolates were avirulent to 11 lines, carrying high-effective *Lr* genes: *9*, *15*, *19*, *24*, *29*, *38*, *41*, *42*, *43*, *47*, *W*.

The cultivar Shef showed a susceptible reaction to all isolates, with the exception of HHTG and PHRT.

1	Isolates										Field damage	
<i>Lr</i> gene	MCPB	FGTD	CGPD	TGPD	PDLG	THRS	PBSS	LHLH	HHTG	PHRT	- Field damage 80S	
1	3	0	2	3	3	3	3	3	2	3		
2a	1	0	0	3	0	3	0	0	3	0	50MS	
2 <i>c</i>	3	3	1	3	3	3	3	2	2	3	60S	
3	3	3	3	3	3	3	3	2	3	3	80S	
3bg	1	0	3	3	3	3	3	2	3	2	90S	
3ka	3	3	3	0	3	3	3	3	3	3	70MS	
9	0	0	0	0	0	0	0	0	0	0	0	
10	0	1	1	0	3	3	2	0	3	3	90S	
11	1	3	1	0	3	3	3	0	3	3	80S	
14a	0	3	3	0	2	3	3	0	0	3	40S	
14b	3	2	2	1	3	3	3	3	3	3	60MS	
15	1	1	1	1	0	1	1	0	1	1	20MS	
16	1	3	3	3	3	3	1	3	3	3	60MS	
17	3	3	3	3	3	0	3	2	3	0	10MR	
18	1	3	0	0	0	0	0	0	0	3	1MR	
19	0	0	0	0	0	0	0	0	0	0	1R	
20	0	1	0	0	2	3	0	1	3	3	30MS	
21	1	1	1	1	1	3	2	3	3	2	15MR	
23	3	3	3	3	3	3	2	3	3	3	30MS	
24	1	0	0	0	0	2	1	0	2	1	R	
25	0	1	1	1	3	3	2	3	2	1	20MR	
26	3	0	0	0	3	3	0	3	3	3	70MS	
28	3	2	0	0	3	3	0	0	3	2	10R	
29	0	0	1	0	1	0	0	1	0	0	R	
80	3	3	3	2	3	3	2	0	3	3	50S	
32	1	1	3	1	1	1	0	0	1	2	20MR	
33	1	3	3	1	2	3	3	3	2	1	40S	
34	2	1	2	3	3	3	1	0	0	2	50S	
86	1	1	3	2	1	3	1	3	3	3	5R	
88	0	0	1	1	0	1	0	1	2	1	10R	
40	3	3	3	3	3	3	2	3	3	1	30MS	
41	1	2	2	0	0	0	0	2	0	0	10R	
12	0	0	0	1	0	1	0	1	0	0	0	
43	0	0	0	0	0	1	0	0	0	0	0	
14	2	1	3	3	3	3	0	3	1	3	20MR	
15	0	0	0	3	0	0	0	0	1	1	5R	
47	0	0	0	0	0	0	0	0	0	0	R	
3	0	1	0	3	2	3	1	2	0	3	60S	
N	1	1	1	0	1	1	1	1	0	1	10MR	
Exch	1	1	0	0	0	3	0	1	3	2	50S	
Kanr	1	3	1	1	1	3	0	0	3	3	60S	
50	0	2	3	1	0	2	0	2	3	1	0	

Table 2. Seedling reaction of near isogenic wheat lines (Lr) on 10 Puccinia triticina isolates (climate chamber, 2017)

S = large uredinia, MS = moderate size uredinia with chlorosis, R = small uredinia with necrosis, MR = moderate size uredinia with necrosis, 0 = no flecks or uredinia

Cultivar	MCPB	FGTD	CGPD	TGPD	PDLG	THRS	PBSS	LHLH	HHTG	PHRT	Field damage
Argonavt	0	2	3	3	3	3	3	0	3	3	20MS
Wintergold	2	3	3	3	3	3	2	0	0	2	10 MR
Yakhont	1	2	3	3	3	0	0	2	2	2	10MS
Etude	3	0	3	3	3	3	1	3	3	3	5MS
Shef	3	3	3	3	3	3	3	3	2	2	10R
Yubilyarka	1	3	3	3	2	3	3	3	2	1	5R
Olkhon	0	0	0	0	0	0	1	0	0	3	1R
Eremeevna	1	0	1	1	3	3	2	3	3	3	5R
Antonina	3	0	3	2	3	3	1	0	3	3	5MR
Anka	2	3	2	1	1	3	0	0	3	1	10R
Krucha	3	3	3	3	3	3	3	0	3	2	5MS
Odari	3	3	3	3	1	3	3	3	0	3	5R
Bagira	1	2	2	0	3	3	0	0	0	0	10MS
Anisimovka	0	2	1	0	0	1	0	0	0	0	10MS
Eirena	3	3	3	0	3	3	3	3	1	3	30MS
Tulaykovskaya 110	0	0	1	0	0	0	2	0	0	0	30MS
Bogdanka	1	3	2	1	2	0	1	0	0	0	0
Vidrada	3	3	3	3	2	3	3	0	3	1	5R
Sidor Kovpak	3	3	3	3	3	3	3	0	2	3	1R
Ekada 113	0	0	0	0	0	0	3	0	0	0	10MR

Table 3. Seedling reaction of 20 winter wheat cultivars, inoculated with 10 virulent Puccinia triticina isolates (climate chamber, 2017)

MS = moderate size uredinia with chlorosis, R = small uredinia with necrosis, MR = moderate size uredinia with necrosis, 0 = no flecks or uredinia

A similar reaction to isolates was observed in Lr2c, which showed low IT (2) to HHTG and high IT (3) to most isolates. Lr40 also exhibited avirulence to PHRT in combination with susceptibility to most *P. triticina* isolates. This suggests the presence of Lr2c and Lr40 in Shef.

The cultivar Krucha showed a stable response to LHLH and PHRT isolates (IT 0, 2), and it was susceptible to other isolates. Resistance to LHLH and susceptibility to other isolates was observed in the line with the Lr3 gene. Low IT (2, 0) to LHLH and PHRT and high IT (3) to most isolates were shown by the line with the Lr17 gene. A similar reaction of the cultivar and these lines makes it possible to postulate the Lr3 and Lr17 genes in Krucha.

Differentiating isolates were not found for the cultivar Anisimovka and Tulaykovskaya 110. Presumably they may contain *Lr* genes: *9*, *29*, *42*, *43*, *47*.

Cultivar Argonavt had a low IT (0, 2) for MCPB, FGTD and LHLH isolates and a high IT (3) for CGPD, TGPD, PDLG, THRS, PBSS, HHTG and PHRT isolates. The MCPB, FGTD, and LHLH isolates were avirulent to the line with the *Lr3bg* gene and virulent to the remaining isolates, with the exception of PHRT, which suggests the presence of *Lr3bg* in Argonavt. PHRT was virulent to *Lr10*, and MCPB. FGTD and LHLH were avirulent to it. Argonavt probably also contains *Lr10*. The PDLG, THRS, LHLH, HHTG, and PHRT isolates showed virulence for the cultivar Eremeevna, and a stable reaction was observed for the isolates MCPB, FGTD, CGPD, TGPD, and PBSS (IT 0, 1, 2). The PDLG, THRS, HHTG, and PHRT isolates were also virulent to *Lr10*, which suggests the presence of this gene in the cultivar Eremeevna. The FGTD, CGPD, TGPD, and PBSS isolates were avirulent to *Lr26*; therefore, Eremeevna may also contain *Lr26*.

Cultivar Antonina showed a stable response to the isolates FGTD, TGPD, PBSS and LHLH, and the rest of the isolates were susceptible. These isolates were avirulent to *Lr10*, which makes it possible to postulate this gene in Antonina. The TGPD, PBSS, and LHLH isolates showed avirulence to *Lr30*, while MCPB, CGPD, PDLG, THRS, HHTG, and PHRT were virulent to it. Perhaps *Lr30* is also present in Antonina.

Odari showed low IT (0.1) for PDLG and HHTG isolates and high IT (3) for other isolates. Isolates PDLG and HHTG showed avirulence to Lr10 and Lr14a, which were susceptible to most other isolates. This allows us to postulate Lr10 and Lr14a in the cultivar.

Olkhon was susceptible only to the PHRT isolate, and showed a stable reaction to the others (IT 0, 1). A combination of virulence to PHRT and resistance to most other isolates, with the exception of FGTD, was observed in *Lr18*. Olkhon, which probably contains *Lr18*. FGTD, and like most isolates, was resistant to *LrB*, and PHRT was virulent to it. Probably, cultivar Olkhon also contains *LrB*.

The cultivar Wintergold showed high IT (3) for FGTD, CGPD, TGPD, PDLG, THRS isolates and low IT (0.2) for MCPB, PBSS, LHLH, HHTG and PHRT. The MCPB and PBSS isolates were avirulent to *Lr16*, while FGTD, CGPD, TGPD, PDLG, and THRS were virulent to it. Wintergold may contain *Lr16*. The PBSS, PHRT, and HHTG isolates showed avirulence to *Lr44*, while CGPD, TGPD, PDLG, and THRS were virulent to the line with this gene. This also makes it possible to postulate *Lr44* in Wintergold. The MCPB, LHLH, and PHRT isolates were avirulent to *Lr3bg*, while CGPD, TGPD, PDLG, and THRS were virulent to the line with this gene, which allows us to postulate *Lr3bg* in the Wintergold cultivar.

Yakhont was resistant (IT 0, 1, 2) to isolates MCPB, FGTD, THRS, PBSS, LHLH, HHTG and PHRT and susceptible (IT 3) to CGPD, TGPD and PDLG. The CGPD, TGPD and PDLG were also susceptible to *Lr44*, while isolates MCPB, FGTD, THRS, PBSS, LHLH and PHRT were resistant to *Lr50*, which allows us to postulate *Lr44* and *Lr50* in Yakhont.

The cultivar Etude showed low IT (0.1) for FGTD and PBSS and high IT (3) for other isolates. PBSS was avirulent to Lr23, and the remaining isolates were virulent to the line with this gene. Therefore, Lr23 can be postulated in Etude. FGTD and PBSS were also avirulent to Lr44, and most of the remaining isolates were virulent to it. Perhaps the cultivar also contains Lr44.

The cultivar Yubilyarka showed stable response (IT 1, 2) to MCPB, PDLG, HHTG and PHRT and was susceptible (IT 3) to other isolates. MCPB, PDLG, and HHTG were avirulent to Lr14a, and isolates FGTD, CGPD, THRS, PBSS, and HHTG were virulent to Lr14a. It is possible that Yubilyarka contains Lr14a. MCPB, PDLG, HHTG, and PHRT were also avirulent to Lr33, and the FGTD, CGPD, THRS, PBSS, and LHLH isolates caused a susceptible response in this line. Therefore, the presence of Lr33 can be assumed in Yubilyarka.

The cultivar Anka showed resistance to most isolates (MCPB, CGPD, TGPD, PDLG, PBSS, LHLH and PHRT) and virulence to FGTD, THRS and HHTG. A similar reaction of isolates was observed in lines with the resistance genes *LrExch* and *LrKanred*. It is likely that the Anka contains these genes.

Bagira also showed a steady response to most isolates, and only PDLG and THRS were virulent. PDLG and THRS which were virulent to Lr25 and most others, with the exception of LHLH, were resistant to this line. This gives reason to postulate Lr25 in the cultivar. LHLH, similar to most isolates, was resistant to Lr10, while PDLG and THRS were susceptible to it. Therefore, Bagira may also contain *Lr10*.

The cultivar Eirena showed a susceptible response to most isolates, with the exception of TGPD and HHTG. TGPD was resistant to Lr3ka, and the remaining isolates were virulent to it. Therefore, the presence of Lr3ka in the cultivar is likely. HHTG was avirulent to Lr2c, and most of the remaining isolates were virulent to the line with this gene. It is possible that Lr2c is also present in Eirena.

Cultivars Bogdanka and Ekada 113 were resistant to most isolates, with the exception of FGTD, which was virulent to Bogdanka, and PBSS, virulent to Ekada 113. Perhaps their resistance was due to the presence of one or more highly resistant *Lr* genes: *9*, *24*, *29*, *38*, *41*, *42*, *43*, *47*, *W*.

The cultivar Vidrada showed low IT (0, 1, 2) for PDLG, LHLH and PHRT isolates and high IT (3) for other isolates. LHLH was avirulent to *Lr3*, which was susceptible to most other isolates. Probably Vidrada contains *Lr3*. LHLH and PHRT were avirulent to *Lr17*, and to the remaining *Lr17* isolates, they were mostly susceptible. Therefore, the cultivar may also contain *Lr17*.

Sidor Kovpak was susceptible to most *P. triticina* isolates, with the exception of LHLH (IT 0) and HHTG (IT 2). LHLH and HHTG were avirulent to *Lr2c*, and the remaining isolates were usually virulent to this gene. This makes it possible to postulate *Lr2c* in Sidor Kovpak.

Adult plant resistance

In the field, the cultivar Bogdanka showed absolute resistance, and in the juvenile phase only one isolate was found that was virulent to this cultivar. Most likely, this cultivar is protected by effective genes of race-specific and age-related resistance.

Six cultivars (Yubilyarka, Olkhon, Yeremeyevna, Odari, Vidrada and Sidor Kovpak) showed high adult resistance (1R – 5R).

Five cultivars (Wintergold, Shef, Antonina, Anka, Ekada 113) showed moderate resistance in the field (5MR, 10R, 10MR). In this case, only Ecada 113 possessed effective genes for juvenile resistance, which work in adulthood as well. In the cultivar Anka, which was resistant to most isolates, *LrExch* and *LrKanred*, not effective in the field, were postulated. Perhaps the resistance of the cultivar was due to the genes for age resistance or their combination. Cultivars Wintergold, Shef and Antonina, which were susceptible to most isolates, are thought to have field ineffective seedling *Lr* genes: *2c*, *3bg*, *10*, *16*, *30*, *40* (30MS – 90S).

The group of cultivars Yakhont, Etude, Krucha, Bagira, Anisimovka, despite a low percentage of the degree of disease damage (5–10%), had a susceptible type of reaction in the field (MS – moderate size of uredinia with chlorosis), which may indicate the effect of gene pyramiding. Differentiating isolates were not found for Anisimovka, which indicates the presence of resistance genes that are effective in the adult stage as well, which nevertheless begin to lose their effectiveness (this is indicated by the susceptible type of reaction). A similar reaction in the field was shown by most of the lines with the genes *Lr17*, *Lr23*, *Lr25*, , *Lr44*, which are postulated in the cultivars Yakhont, Etude, Krucha and Bagira.

Cultivars Argonavt, Eirena and Tulaykovskaya 110 in the field were moderately susceptible to *P. triticina* (20S – 30S), but have a different genetic basis. Differentiating isolates were not found for Tulaykovskaya 110, which may indicate the presence of one or more genes from among 11 lines with *Lr* genes, to which no isolates were found.

In cultivars Argonavt and Eirena, field-ineffective seedling genes *Lr2c*, *Lr3ka*, *Lr3bg*, and *Lr10* were postulated.

Discussion

In 20 winter wheat cultivars from different breeding centers of Russia, Ukraine and Germany, 19 resistance genes *Lr* were postulated: *2c*, *3*, *10*, *3bg*, *3ka*, *14a*, *17*, *18*, *23*, *25*, *26*, *30*, *33*, *40*, *44*, *50*, *B*, *Exch*, *Kanred*. The *Lr10* gene was the most common in cultivars (Argonavt, Antonina, Eremeevna, Odari, Bagira). Long-term studies of E.I. Gultyaeva (Gultyaeva 2018) confirm, with the help of molecular markers, the wide distribution of the ineffective seedling *Lr10* gene in Russian cultivars. The presence of *Lr10* in the cultivars Bagira and Antonina is also confirmed by it with the help of molecular markers.

For two cultivars (Tulaykovskaya 110 and Anisimovka) no differentiating isolates were found. Moreover, in the field, cultivars were moderately susceptible, with a low degree of damage, which may indicate the presence of several additional genes that give the effect of pyramiding. Tulaykovskaya 110 was developed with the participation of *Elytrigia intermedia* (wheatgrass intermediate), from which the effective *LrAg* gene was introgressed into soft wheat (Sochalova and Lichenko 2013).

The cultivar Bogdanka was the only studied cultivar that showed absolute resistance in the field and low IT (0, 1, 2) to all isolates except FGTD. According to Kozub *et al.* (2012), this cultivar has rye translocation on chromosome 1A (1AL/1RS), which is rare for Russian varieties. Soft wheat cultivars carrying the wheat rye translocation 1AL/1RS contains a combination of genes effective against several diseases (Crespo-Herrera *et al.* 2017).

Cultivars Yubilyarka, Yeremeyevna, Odari, Vidrada and Sidor Kovpak, which demonstrated high field resistance, showed a high IT (3) to most isolates in the seedling phase. Most seedling genes postulated in these cultivars (Lr2c, Lr3, Lr 10, Lr 14a, Lr 26, Lr 33) were ineffective in the field (40S – 90S). In the cultivar Olkhon, Lr18 provided resistance in the seedling phase, which was also effective in the adult stage. Resistance of the remaining cultivars may be associated with adult plant resistance genes.

Most of the studied cultivars have recently been entered into the State Register of the Russian Federation (2015–2019) and in the field show a stable or moderately susceptible reaction, despite the fact that the Lr genes postulated in them are not effective in the adult stage of plants. Therefore, annual monitoring of the resistance of cultivars is required, especially with a medium susceptible type of reaction (MS), since this indicates the initial stage of resistance loss.

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References

- Abouzied H., Argawy E., Orabey E. 2017. Molecular markers and postulation study of leaf rust resistance genes in various Egyptian wheat cultivars. Biotechnology Journal International 20 (2): 1–13. DOI: https://doi.org/10.9734/ BJI/2017/36175
- Anpilogova L., Volkova G. 2000. Methods for creating artificial infectious backgrounds and assessing wheat cultivars for resistance to harmful diseases (spike fusarium, rust, powdery mildew): guidelines. Krasnodar, 28 pp.
- Anpilogova L., Volkova G., Vaganova O., Avdeeva Yu. 2011. Identification of genes of juvenile resistance to the leaf rust pathogen of in new domestic winter wheat cultivars. Plant Protection Newsletter 3: 38–40.
- Baidya S., Bhardwaj S., Shrestha S., Manandhar H., Thapa R., Mahto N., Joshi A. 2019. Evaluation of wheat genotypes for seedling and adult plant resistance to leaf rust (*Puccinia triticina*). Discovery Agriculture 5: 69–78.
- Crespo-Herrera L.A., Garkava-Gustavsson L., Åhman I. 2017. A systematic review of rye (*Secale cereale* L.) as a source of

resistance to pathogens and pests in wheat (*Triticum aestivum* L.). Hereditas 154: 11. DOI: 10.1186/s41065-017-0033-5

- Flor H. 1971. Current status of the gene-for-gene concept. Annual Review of Phytopathology 9: 275–296.
- Gebrewahid T., Yao Z., Yan X., Gao P., Li. Z. 2017. Identification of leaf rust resistance genesin Chinese common wheat cultivars. Plant Disease 101: 1729–1737. DOI: https://doi. org/10.1094/PDIS-02-17-0247-RE
- Gultyaeva E. 2018. Genetic structure of *Puccinia triticina* populations in Russia and its variability under the influence of the host plant. Ph.D. Abstract thesis, Saint Petersburg-Puszkin, 42 pp.
- Kozub N., Sozinov I., Sobko T., Dedkova O., Badaeva E., Netsvetaev V. 2012. Rye translocation in some soft winter wheat cultivars. Agricultural Biology 3: 68–74. DOI: https://doi. org/10.15389/agrobiology.2012.3.68rus
- Li Z., Xia X., He Z., Li X., Zhang L., Wang H., Meng Q., Yang W., Li G., Liu D. 2010. Seedling and slow rusting resistance to leaf rust in Chinese wheat cultivars. Plant Disease 94: 45–53. DOI: https://doi.org/10.1094/PDIS-94-1-0045
- Li T., Wu X., Xu X., Wang W., Cao Y. 2016. Postulation of seedling stem rust resistance genes of Yunnan wheat cultivars in China. Plant Protection Science 52: 242–249. DOI: https:// doi.org/10.17221/137/2015-PPS
- Li J., Shi L., Wang X., Zhang N., Wei X., Zhang L., Yang W., Liu D. 2018. Leaf rust resistance of 35 wheat cultivars (lines). Journal of Plant Pathology and Microbiology 9 (1): 429. DOI: https://doi.org/10.4172/2157-7471.1000429
- Long D., Kolmer J. 1989. North American system of nomenclature for *Puccinia recondita* f. sp. *tritici*. Phytopatology 79: 525–529.
- Mebrate S., Dehne H., Pillen K., Oerke E. 2008. Postulation of seedling leaf rust resistance genes in selected Ethiopian and German bread wheat cultivars. Crop Science 48: 507–516. DOI: https://doi.org/10.2135/cropsci2007.03.0173
- Oelke L., Kolmer J. 2004. Characterization of leaf rust resistance in hard red spring wheat cultivars. Plant Disease 88: 1127–1133. DOI: https://doi.org/2004-0727-02R

- Sanin S., Nazarova A. 2010. Phytosanitary situation on wheat crops in the Russian Federation (1991–2008). Analytical Review. Protection and Quarantine of Plants 2: 70–78.
- Shcherbik A., Kovalenko E. 2011. Selection of donors of wheat resistance to leaf rust. Protection and Quarantine of Plants 2: 45–46.
- Sochalova L., Lichenko I. 2013. Gene pool of resistance sources of soft spring wheat to leaf-stem diseases. Achievements of Science and Technology of the Agro-Industrial Complex 6. Available on: https://cyberleninka.ru/article/n/genofondistochnikov-ustoychivosti-myagkoy-yarovoy-pshenitsy-klistosteblevym-zabolevaniyam [Accessed: 20 January 2020]
- Vanzetti L., Campos P., Demichelis M., Lombardo L., Aurelia P., Vaschetto L., Bainotti C., Helguera M. 2011. Identification of leaf rust resistance genes in selected Argentinean bread wheat cultivars by gene postulation and molecular markers. Electronic Journal of Biotechnology 14 (3): 9. DOI: https:// doi.org/10.2225/vol14-issue3-fulltext-14
- Volkova G., Kudinova O., Vaganova O. 2019. Screening of wheat Lr genes for resistance to Puccinia triticina in the North Caucasus region. Bulletin of the Russian Agricultural Science 5: 54–56. DOI: https://doi.org/10.30850/vrsn/2019/5/54-56
- Volkova G., Vaganova O. 2016. Postulation of leaf rust resistance genes in cultivars of soft winter wheat. Journal of International Scientific Publications: Agriculture & Food 4: 627–632.
- Wamishe Y., Thompson K., Milus E. 2004. A computer program to improve efficiency and accuracy of postulating racespecific resistance genes. Plant Disease 88: 545–549. DOI: https://doi.org/10.1094/PDIS.2004.88.5.545
- Wamishe Y., Milus E. 2004. Seedling resistance genes to leaf rust in soft red winter wheat. Plant Disease 88: 136–146. DOI: https://doi.org/10.1094/PDIS.2004.88.2.136
- Wei X., Zhang H., Du D., Yang W., Liu D. 2015. Evaluation of wheat leaf rust resistance genes in 10 wheat genotypes. Journal of Plant Diseases and Protection 122 (2): 91–99. DOI: https://doi.org/10.1007/BF03356536